

## In Vitro pO<sub>2</sub> Measurement of Islet Encapsulation Devices in Oxygen Measurement Core

Mignavani Kotecha<sup>1\*</sup>, Zhengshan Zhao<sup>1</sup>, Darwin Boderio<sup>1</sup>, Eliyas Siddiqui<sup>1</sup>, Marcelo Gutierrez<sup>1</sup>, Jovan Mitrevski<sup>1</sup>, Longhai Wang<sup>2</sup>, Alexander Ulrich Ernst<sup>2</sup>, Lisa Danielczak<sup>3</sup>, Stephanie Fernandez<sup>3</sup>, Minglin Ma<sup>2</sup>, Corinne Hoesli<sup>3</sup>, Cherie Stabler<sup>4</sup>, Klearchos Papas<sup>5</sup>, and Boris Epel<sup>6</sup>

<sup>1</sup>Oxygen Measurement core, O2M Technologies, 2242, W. Harrison Street, STE 201-18, Chicago, IL, USA. <sup>2</sup>Dept of Biological and Environmental Engineering, Cornell University. <sup>3</sup>Department of Chemical Engineering, McGill University. <sup>4</sup>Department of Biomedical Engineering, University of Florida. <sup>5</sup>Department of Surgery, University of Arizona. <sup>6</sup>Department of Radiation and Cellular Oncology, The University of Chicago.

\*Contact: mkotecha@o2map.com

**Statement of Purpose:** The lack of oxygen supply to the highly metabolic pancreatic islet cells is one of the major factors contributing to the failure of islet transplantation devices targeting the cure of type I diabetes (T1D). The loss of islets due to hypoxia is common in almost all modes of islet transplantation- micro-encapsulation devices, macro-encapsulation devices, and tissue-grafts transplantation. Several approaches to improve oxygenation in these transplantation devices are thus being tested (1-4). However, because of the lack of available technologies to provide oxygen partial pressure (pO<sub>2</sub>) assessment in and around devices, the progress is severely hindered.

**O2M Technologies' platform non-invasive Oxygen Imaging technology** has the potential to guide the development of islet cell transplantation therapies by providing real-time high accuracy pre- and post-implantation pO<sub>2</sub> maps in and around devices *in vitro* and *in vivo* (5-6). O2M's preclinical small animal oxygen imager, JIVA-25 (Figure 1A), provides average pO<sub>2</sub> values in sample volumes (up to 40 mm) as well as three-dimensional pO<sub>2</sub> maps with high spatial (0.5 mm, isotropic), temporal (1-10 min), and pO<sub>2</sub> (1-3 torr) resolution. For reporting oxygen concentration, JIVA-25 uses oxygen-dependent relaxation rates of trityl radicals OX063 or its deuterated version OX071 (Figure 1B). The pO<sub>2</sub> is linearly related to R<sub>1</sub> (Figure 1C). JIVA-25 uses spin-echo based T<sub>1</sub> inversion recovery method for mapping absolute pO<sub>2</sub>. For the measurements, samples can be loaded vertically or horizontally (Figure 1D-1F).

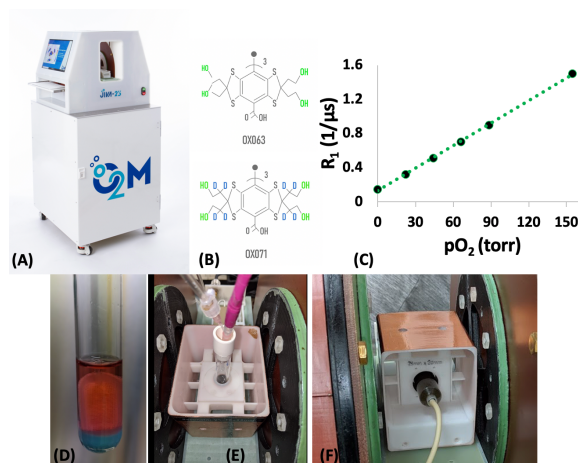


Figure 1: (A) JIVA-25 Instrument. (B) hydroxyethyl tetrathiatrityl methyl radical OX063 and OX071, (C) Example calibration curve at 25 mT, the oxygen in the sample (PBS at 37 °C) was set at 0, 3, 6, 9, 12, and 21% and R<sub>1</sub> were measured when the equilibrium was reached. (D) A 20 μL TheraCyt device in a 16 mm tube, (E) Example *in vitro* measurement set up for vertical access resonator, (F) example *in vitro* measurement for horizontal access resonators inside JIVA-25.

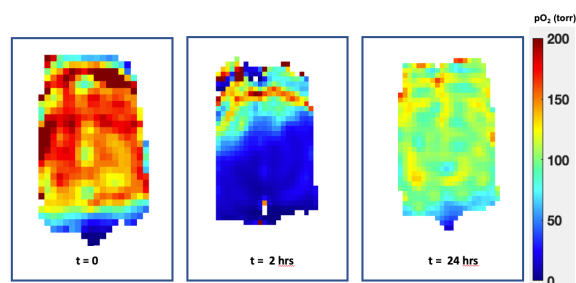


Figure 2: pO<sub>2</sub> maps of 10 million beta TC6 cells loaded 20 μL TheraCyt device in a 16 mm test tube at different time points during a 24-hour experiment. The incubator gas mixture (95% air and 5% CO<sub>2</sub>) was circulated on top of the sample throughout the experiment. The temp was kept at 37 (± 1) °C.

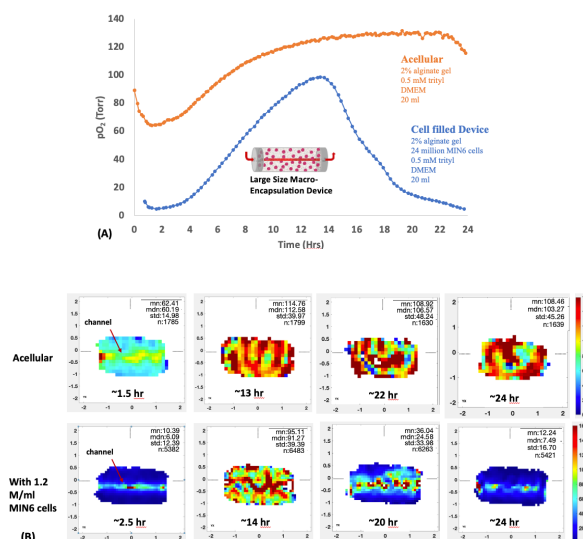


Figure 3: (A) Average pO<sub>2</sub> and (B) pO<sub>2</sub> maps and statistics of a large size macro-encapsulation device with a perfusion channel in the middle and filled with either only 2% alginate (Acellular) or with 24 million MIN6 cells embedded in 2% alginate as a function of time for a 24-hour experiment. The trityl concentration was 0.5 mM in the sample and 0.25 mM in the channel.

This work is the outcome of JDRF-supported “Oxygen Measurement Core” facility established at O2M Technologies in 2019. We performed *in vitro* pO<sub>2</sub> measurements of acellular and cell loaded islet cell transplantation devices at the core. These devices varied in shape, size, biomaterials, and oxygen profile. We will present key data from these measurements. Some example data are presented in Figure 2 and 3.

**Acknowledgements:** JDRF 3-SRA-2020-833-M-B, NIH R43CA224840, R44CA224840, NSF 1819583.

**References:** 1. Bowers et al., Acta Biomater. 95:131-151, 2019. 2. Coronel et al., Biomaterials, 129:139-151, 2017. 3. Fernandez et al., Front. Bioeng. Biotechnol. Conference abstract:10<sup>th</sup> world biomaterial congress 2016. 4. Papas et al., Adv Drug Deliv Rev. 139:139-156, 2019. 5. Epel et al., J. Mag Reson. 280:149-157, 2017. 6. Kotecha et al., Tissue Eng Part C Methods, 24(1):14-19, 2018.